

Crystal and molecular structure of allyl *O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosylonate)-(2 $\rightarrow$ 8)-*O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosidonate)-monohydrate

Vincent Mikol ^{a,*}, Paul Kosma ^b, Helmut Brade ^c

^a Preclinical Research, Sandoz Pharma, CH-4002 Basel Switzerland

^b Institut für Chemie der Universität für Bodenkultur Wien, A-1180 Wien Austria

^c Forschungsinstitut Borstel, D-23845 Borstel Germany

Received 14 March 1994; accepted 6 May 1994

Abstract

The α -(2 $\rightarrow$ 8)-linked Kdo disaccharide derivative allyl *O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosylonate)-(2 $\rightarrow$ 8)-*O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosidonate)-monohydrate $C_{19}H_{28}O_{15}Na_2 \cdot H_2O$, $M_r = 542.32$, is orthorhombic, $P2_12_12_1$ with $a = 9.229(1)$, $b = 12.036(1)$, $c = 21.671(1)$ Å, and $Z = 4$. The structure was solved by direct methods and refined to $R = 0.040$ for 2677 observed reflections. The torsion angles about the (2 $\rightarrow$ 8)-glycosidic bond are stabilized by an intramolecular hydrogen bond between the carboxylate group at the anomeric carbon atom of the terminal Kdo residue and the hydroxyl group O-17 of the second Kdo moiety.

Keywords: Chlamydia; Disaccharide; Kdo ; α -(2 $\rightarrow$ 8)-Glycosidic linkage; Crystal structure

1. Introduction

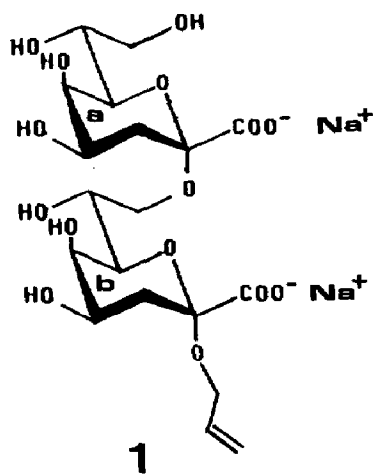
Chlamydiae are pathogenic, obligatory intracellular parasites which cause diseases in animals and humans [1]. They possess, in their outer membrane, a lipopolysaccharide which constitutes one of the major surface antigens of Chlamydia harboring a carbohydrate epitope which is shared by the whole genus. The epitope is composed of a trisaccharide of 3-deoxy-D-manno-2-octulosonic acid (Kdo) of the sequence α -Kdop-(2 $\rightarrow$ 8)- α -Kdop-

* Corresponding author.

(2 → 4)-Kdo in which the (2 → 8)-linked disaccharide portion represents the immunodominant part [2]. The primary structure of the Kdo trisaccharide has been determined [3], it has been chemically synthesized [4], and monoclonal antibodies of high affinity have been prepared against it [5]. However, only preliminary data have been obtained on the conformation of this unique carbohydrate antigen [6]. We report here on the crystal structure of a Kdo disaccharide and provide new data on the stereochemistry of the α -(2 → 8)-glycosidic bonding.

2. Experimental

The α -(2 → 8)-linked Kdo disaccharide derivative **1** was synthesized as previously described [4]. Suitable crystals were obtained from aq EtOH at room temperature by adding



dry EtOH in portions during several weeks to a solution of **1** (63 mg) in H₂O (0.3 mL) until a final volume of 1.5 mL was reached; mp 284–287°C (dec). Anal. Calcd for C₁₉H₂₈Na₂O₁₅ · H₂O: C, 40.72; H, 5.39. Found: C, 40.20; H, 5.23. The crystal data and details of the intensity-data collection and structure determination are given in Table 1. The data were not corrected for absorption effect. The structure was solved with SHELX76 [7], and refined anisotropically for non-H atoms. All H atoms were located in difference Fourier maps and refined with isotropic thermal parameters. The final positional parameters are given in Table, the description of the internal geometry of the (2 → 8)-glycosidic linkage is shown in Table 3, and the co-ordination of the sodium ions and the hydrogen bond geometry are given in Tables 4 and 5, respectively. Fig. 1. shows the chemical structure and numbering of atoms.

3. Results and discussion

The conformation of the molecule is shown in Fig. 1. A least square superposition of the ammonium Kdo [8] pyranose ring atoms on those of **1** gives a root-mean-square difference

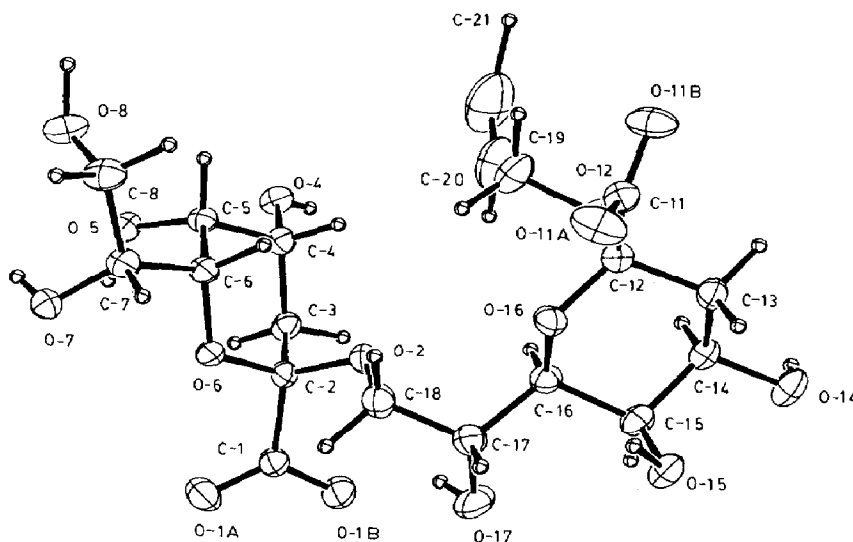


Fig. 1. Perspective view and atom labelling of allyl *O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosylonate)-(2 $\rightarrow$ 8)-*O*-(sodium 3-deoxy- α -D-manno-2-octulopyranosidonate)-monohydrate. The thermal ellipsoids are at 50% probability. H-atoms are represented by spheres of arbitrary size.

of 0.023 and of 0.028 Å for the atoms of units **a** and **b**, respectively. This indicates that both pyranose rings adopt a 5C_2 chair conformation as already noticed for a methyl ketoside of Kdo [9]. The ring pucker places the carboxylate groups in an equatorial position. Including the non-hydrogen axial and equatorial substituents of ammonium Kdo, the superposition on unit **a** and **b** yields a root-mean-square difference of 0.084 and 0.068 Å for unit **a** and **b**, respectively, suggesting a rather identical conformation. The two most significant variations for units **a** and **b** are: (i) similar to the methyl ketoside of Kdo [9], the C-1–O-1B bond is synperiplanar to C-2–O-2 whereas in the case of ammonium Kdo the carboxyl bond nearly eclipses the C-2–O-2 bond. This was thought to be a likely result of the presence of a

Table 1

Crystallographic Data for **1** (Enraf–Nonius CAD4 diffractometer)

Crystal dimensions (mm)	0.1 $\times$ 0.2 $\times$ 0.2
Space group	$P2_12_12_1$
Cell dimensions (Å) <i>a</i> , <i>b</i> , <i>c</i>	9.229(1), 12.036(1), 21.671(1)
Cell volume Å ³	2406.43
<i>Z</i>	4
<i>F</i> (000)	1136
Calculated density (g/cm ⁻³)	1.5
λ , CuK α (Å)	1.5406
μ (cm ⁻¹)	13.83524
$2\theta_{\max}$ (°)	148.6
Number of reflections measured	2783
Number of reflections observed ($F_o > 4\sigma(F_o)$)	2677
Number of refined parameters	373
Final residual factors: <i>R</i> , <i>R_w</i>	0.0401, 0.0462

Table 2

Fractional positional parameters ($\times 10^4$) and equivalent isotropic displacement coefficients ($\text{\AA}^2$) for non-H atoms (U_{eq}) and isotropic displacement coefficient for H-atoms (U_{iso}). The standard deviations are given in parentheses. $U_{\text{eq}} = 1/3 \sum_i \sum_j U_{ij} a_i^* a_j^* a_i a_j$

Atom	x	y	z	$U_{\text{eq}}/U_{\text{iso}} (\times 10^3)$
C-1	−2224(3)	6645(2)	3680(1)	25(1)
O-1A	−1777(2)	7381(2)	3334(1)	43(1)
O-1B	−3456(2)	6618(2)	3934(1)	34(1)
C-2	−1245(1)	5612(2)	3799(1)	21(1)
O-2	−1256(2)	5304(1)	4430(1)	26(1)
C-3	−1805(1)	4601(2)	3442(1)	25(1)
H-3A	−1977(3)	4832(2)	2966(1)	50(1)
H-3B	−2818(3)	4333(2)	3642(1)	50(1)
C-4	−721(1)	3645(2)	3470(1)	22(1)
H-4A	−670(3)	3389(2)	3947(1)	51(1)
O-4	−1172(2)	2727(2)	3103(1)	28(1)
H-4	−1900(60)	2520(40)	3175(22)	62(2)
C-5	779(3)	4015(2)	3256(1)	21(1)
H-5A	1567(3)	3368(2)	3337(1)	51(1)
O-5	844(2)	4178(2)	2605(1)	26(1)
H-5	380(40)	488(30)	2473(19)	42(1)
C-6	1218(3)	5031(2)	3638(1)	21(1)
H-6A	1310(3)	4654(2)	4089(1)	52(1)
O-6	167(2)	5902(1)	3597(1)	22(1)
C-7	2660(3)	5594(2)	3490(1)	25(1)
H-7A	2849(3)	6179(2)	3860(1)	45(1)
O-7	2589(2)	6233(2)	2938(1)	28(1)
H-7	2870(50)	5940(30)	2680(19)	43(1)
C-8	3961(3)	4821(2)	3521(2)	33(1)
H-8A	3891(3)	4330(2)	3937(2)	51(1)
H-8B	4939(3)	5314(2)	3531(2)	51(1)
O-8	4017(2)	4098(2)	2006(1)	37(1)
H-8	4680(60)	3400(40)	3045(21)	65(1)
C-11	715(3)	3566(2)	6602(1)	25(1)
O-11A	1498(3)	4370(2)	6735(1)	40(1)
O-11B	1003(2)	2568(2)	6715(1)	37(1)
C-12	−786(3)	3790(2)	6308(1)	23(1)
O-12	−1227(2)	2915(2)	5912(1)	28(1)
C-13	−1933(3)	3816(2)	6820(1)	26(1)
H-13A	−1628(3)	4419(2)	7165(1)	53(1)
H-13B	−2016(3)	3005(2)	7030(1)	53(1)
C-14	−3386(3)	4137(2)	6540(1)	26(1)
H-14A	−3701(3)	3527(2)	6200(1)	51(1)
O-14	−4493(2)	4154(2)	7006(1)	34(1)
H-14	−4940(8)	3620(50)	6860(30)	90(2)
C-15	−3242(3)	5264(2)	6224(1)	26(1)
H-15A	−4256(3)	5507(2)	6015(1)	48(1)
O-15	−2919(2)	6114(2)	6657(1)	30(1)
H-15	−1620(60)	6250(50)	6690(30)	82(2)
C-16	−2079(3)	5153(2)	5723(1)	25(1)
H-16A	−2491(3)	4545(2)	5400(1)	52(1)
O-16	−720(2)	4816(2)	5991(1)	26(1)

Table 2 (continued)

Atom	x	y	z	$U_{eq}/U_{iso} (\times 10^3)$
C-17	−1787(4)	6254(2)	5385(1)	31(1)
H-17A	−1262(4)	6825(2)	5698(1)	47(1)
O-17	−3153(3)	6697(3)	5193(1)	52(1)
H-17	−3240(50)	6390(40)	4820(21)	58(1)
C-18	−725(3)	6138(2)	4856(1)	31(1)
H-18A	319(3)	5889(2)	5033(1)	51(1)
H-18B	−622(3)	6924(2)	4620(1)	51(1)
C-19	−304(4)	2692(3)	5395(1)	42(2)
H-19A	88(4)	3462(3)	5200(1)	52(2)
H-19B	602(4)	2186(3)	5536(1)	52(2)
C-20	−1206(5)	2085(5)	4932(2)	61(2)
H-20A	−2124(5)	2535(5)	4749(2)	52(3)
C-21	−1030(8)	1115(5)	4728(3)	86(4)
H-21A	−1700(60)	810(40)	4349(22)	67(1)
H-21B	80(8)	190(5)	4920(30)	120(30)
Na-1	517(1)	7545(1)	2957(1)	30(1)
Na-2	1902(1)	6131(1)	7010(1)	35(1)
O-1W	204(3)	2156(2)	8181(1)	38(1)
H-1W	−170(60)	2600(40)	8550(23)	73(2)
H-2W	360(50)	2490(30)	7913(20)	46(1)

methyl group on the anomeric carbon [9]; (ii) a different conformation about the C-6–C-7 was found in unit **a** where C-5–C-6 and C-7–C-8 are in the *gauche* orientation, whereas in the structure of ammonium Kdo, as well as in unit **b**, these bonds are *trans* to one another. This is presumably the result of crystal packing contacts. Interestingly, the solution conformation of unit **b**, as indicated by the coupling constant $^3J_{6,7}$ of the disaccharide **1** ($J_{6,7} \approx 9.0$ Hz), is different from that observed with higher oligosaccharides containing the *Chlamydia* specific trisaccharide epitope α -Kdop-(2 $\rightarrow$ 8)- α -Kdop-(2 $\rightarrow$ 4)- α -Kdop ($J_{6,7} \approx 4.0$ –5.1 Hz).

The C-2–O-2–C-18 bond angle is $115.5(2)^\circ$ and the C–O bond distances are $1.417(3)$ and $1.450(3)$ Å, respectively. The striking feature of this glycosidic linkage is the presence of an intramolecular hydrogen bond between one carboxylate atom of the unit **a** and the hydroxyl group (O-17–H-17) of the second unit **b** which donates a proton. The hydrogen atom H-17 lies at a distance of 2.41 Å from the glycosidic oxygen O-2 suggestive of a three-center hydrogen bond [10] (Table 5). This H-bond makes the glycosidic linkage rigid and reduces the flexibility around the O-2–C-18 bond. It places the two pyranose rings at an angle of ca. 60° . As the torsion angles indicate, the C-2–O-2 bond is synclinal to C-18–C-17 and O-2–C-18 is synperiplanar to C-17–C-16.

A stereoscopic drawing of the crystal structure, viewed along the *a*-axis is shown in Fig. 2. All the oxygen atoms, except the ring oxygen O-12, participate in the hydrogen bond network. The structure contains two independent sodium ions. The first one Na1 shows a six-fold oxygen coordination, whereas the second sodium has a five-fold coordination involving a distorted trigonal bipyramide. The Na1–O and Na2–O distances average

Table 3

Selected bond lengths, bond angles, and torsion angles around the (2 → 8)-glycosidic linkage

Bond lengths (Å)	
C-1–C-2	1.558(3)
O-6–C-2	1.419(3)
C-2–O-2	1.417(3)
O-2–C-18	1.450(3)
C-18–C-17	1.515(4)
C-17–C-16	1.537(4)
Bond angles (°)	
O-1B–C-1–C-2	115.4(2)
O-1A–C-1–C-2	118.7(2)
O-6–C-2–O-2	111.6(2)
C-1–C-2–O-2	111.4(2)
C-2–O-2–C-18	115.5(2)
O-2–C-18–C-17	109.1(2)
C-18–C-17–C-16	113.2(2)
Torsion angles (°)	
O-1A–C-1–C-2–O-2	137.6(2)
O-1B–C-1–C-2–O-6	–167.5(2)
O-6–C-2–O-2–C-18	58.1(3)
C-1–C-2–O-2–C-18	–60.8(3)
C-2–O-2–C-18–C-17	131.9(3)
O-2–C-18–C-17–C-16	–56.2(3)
O-2–C-18–C-17–O-17	66.2(3)
C-18–C-17–C-16–C-15	175.6(2)

2.44(12) and 2.34(9) Å, and fall in the expected range of Na–O distances for sodium ions having a co-ordination number of six and five, respectively [11].

The crystal packing, projected along the *a*-axis, is dominated by the co-ordination of the sodium ions and hydrogen bonds involving the water molecule. The Kdo unit **a** has its pyranose ring almost parallel to the *a*–*b* layer.

Table 4

Geometry of the sodium co-ordination. The distances are given in Å. O-1A and O-11A are carboxylate oxygens, O-15, O-4, O-7, O-5, and O-8 are hydroxyl oxygen atoms, O-6 is a ring oxygen, and O-1W the water oxygen. Symmetry code: (I) *x*, *y*, *z*; (II) $-x + 1/2, -y, z + 1/2$; (III) $-x, y + 1/2, -z + 1/2$; (IV) $1/2 + x, -y + 1/2, -z$

Na1...O-1A ^(I)	2.28
Na1...O-15 ^(IV+b+c)	2.32
Na1...O-4 ^(III)	2.39
Na1...O-6 ^(I)	2.44
Na1...O-7 ^(I)	2.48
Na1...O-5 ^(III)	2.63
Na2...O-11A ^(I)	2.23
Na2...O-1A ^(IV+b+c)	2.29
Na2...O-8 ^(II+b)	2.34
Na2...O-1W ^(III+c)	2.34
Na2...O-5 ^(II+b)	2.48

Table 5

Geometry of the hydrogen-bonding pattern and short contacts. The distances are given in Å and the angles in degrees. For the symmetry codes, see Table 4. The weak hydrogen bonds, which are a minor component of the three- or four-center bonds, are listed in the lines below that of the major hydrogen-bond component. All the minor components are intramolecular hydrogen bonds

Donor	Acceptor	$d(\text{D}-\text{H})$	$d(\text{D} \cdots \text{A})$	$d(\text{H} \cdots \text{A})^a$	Angle ($\text{D}-\text{H} \cdots \text{A})^a$
O-4-H-4	O-11B ^(IV-a+c)	0.74	2.66	1.94 (1.74)	162 (159)
O-5-H-5	O-14 ^(II-a+b+c)	0.98	2.70	1.75 (1.76)	160 (159)
	O-7		3.04	2.80 (2.80)	94 (94)
	O-6		3.05	2.74 (2.73)	99 (100)
O-1W-H-1W1	O-1B ^(II-a+b)	1.02	2.73	1.78 (1.84)	152 (153)
O-17-H-17	O-1B	0.89	2.74	1.95 (1.89)	148 (147)
	O-2		2.94	2.41 (2.39)	118 (116)
O-14-H-14	O-4 ^(IV-a+c)	0.83	2.75	1.98 (1.84)	156 (153)
O-8-H-8	O-11B ^(IV+c)	1.04	2.78	1.76 (1.83)	164 (164)
O-15-H-15	O-1W ^(III+c)	1.21	2.82	1.73 (1.93)	148 (152)
	O-16		2.94	2.45 (2.41)	102 (107)
O-7-H-7	O-11A ^(II+b-c)	0.71	2.83	2.16 (1.92)	158 (157)
	O-8		2.89	2.56 (2.49)	111 (105)
	O-5		3.04	2.83 (2.80)	100 (95)
O-1W-H-1W2	O-7 ^(II+b)	0.72	2.86	2.43 (2.34)	120 (113)

^a Values after the D-H distance were normalized to 0.97 Å and are given in parentheses.

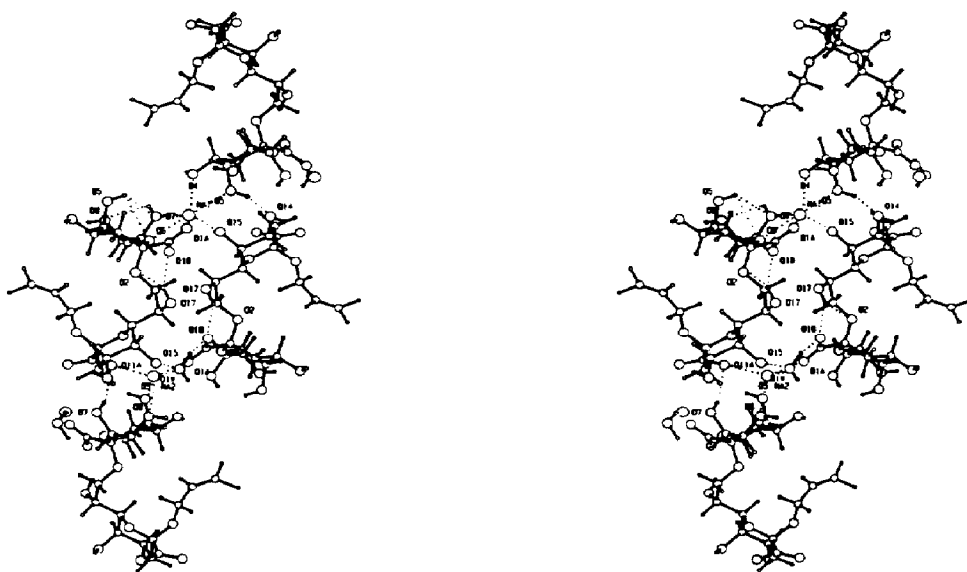


Fig. 2. Stereoview of the unit cell of **1** along the *a* axis (the *c*-axis is vertical). Dotted lines indicate hydrogen bonds or sodium coordination.

Acknowledgements

H.-P. Weber is thanked for help in the structure determination. Financial support for the synthesis of **1** is gratefully acknowledged (Fonds zur Förderung der wissenschaftlichen Forschung-Projekt P 8203 to P.K.).

References

- [1] J. Schachter, *Curr. Top. Microbiol. Immunol.*, 138 (1988) 109–139.
- [2] H. Brade, L. Brade, and F.E. Nano, *Proc. Natl. Acad. Sci. U.S.A.*, 84 (1987) 2508–2512.
- [3] O. Holst, L. Brade, P. Kosma, and H. Brade, *J. Bacteriol.*, 173 (1991) 1862–1866.
- [4] P. Kosma, R. Bahnmüller, G. Schulz, and H. Brade, *Carbohydr. Res.*, 208 (1990) 37–50.
- [5] Y. Fu, M. Baumann, P. Kosma, L. Brade, and H. Brade, *Infect. Immun.*, 60 (1992) 1314–1321.
- [6] K. Bock, J.U. Thomsen, P. Kosma, R. Christian, O. Holst, and H. Brade, *Carbohydr. Res.*, 229 (1992) 213–224.
- [7] G.M. Sheldrick, SHELX76, A Program for Crystal Structure Determination, University of Cambridge, UK, 1976.
- [8] G.I. Birnbaum, R. Roy, J.-R. Brisson, and H.J. Jennings, *J. Carbohydr. Chem.*, 6 (1987) 17–39.
- [9] C. Kratky, D. Stix, and F.M. Unger, *Carbohydr. Res.*, 92 (1981) 299–304.
- [10] G.A. Jeffrey and W. Saenger, *Hydrogen Bonding in Biological Structures*, Springer-Verlag, Berlin, 1991, pp 136–146.
- [11] I.D. Brown, *Acta Crystallogr., Sect. B*, 44 (1988) 545–553.